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the materials for fabricating and testing our microfluidic thermometer are all available from the american institute of bioscience and biotechnology (a.i.b.) and are all listed in 30. we employed a variety of microfluidic thermometers, so we recommend that you read the fabrication and operation sections of this article carefully, rather than using trial and error methods to find your own set-up. note that we have not simulated the effects of melt-crystallization or storage on the freezing point of the mixture of water and ice. we recommend carrying out measurements of samples stored at various temperatures to determine the difference in freezing temperatures with and without storage. to demonstrate how our method performs on a common exercise, we measured the freezing point of a mixture of water and powdered sugar at various volumes. as fig 1 shows, this 3d printed chip has access holes on the bottom that permit the inlet and outlet tubes and fluid connections (blue connectors) to be inserted while the device is sealed on all sides. we loaded the s1 file for this article onto the chip and placed it in a fume hood to cure. we then attached the chip to a slide and carefully placed the chip and tubing in an environmental chamber (spectros, spectronics corporation, il) so that the sample fluid and plastic tubing connections did not contact the chip. the temperature of the environmental chamber was carefully calibrated. we plugged the chip with more tubing and placed it in a temperature-controlled chamber and carefully measured the change in volume of the fluid and change in temperature. we used a digital thermometer to measure the temperature of the unknown solution, attached to a long piece of plastic tubing so that it was outside of the environmental chamber (figure 1). the temperature of the unknown solution was systematically lowered from 20c to -20c and back again. after each temperature change, we extracted the chips, plugged the inlets with plastic tubing, placed the chip back in the environmental chamber, and measured the change in volume.

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each channel was filled with blue food coloring to enhance the visibility of the oil-water interface. at the top of each channel, a solution of nacl (30% solution; fisher scientific) was added to each channel (500 nl for each channel) and left to equilibrate by allowing it to flow out the desired channel at 1 cm/minute and allowed to dry. a 1 ml syringe (becton dickinson) was used to fill the chip, exerting 0.5 n vacuum to eliminate all residual bubbles. the chip was then immediately placed on a clean table and capped with a plastic sheet (without a pressure seal) to prevent the escape of air. for each channel, the solid-liquid interface is clearly visible in the microchannel, and a linear temperature gradient can be established along the lateral length of each channel. to set the temperatures in the channels to zero degrees celsius, a 1 cm-long e.g. platinum heater (vwr international) and a pair of electrodes (made from 0.1-inch and 0.01-inch (awg) stainless steel wire, respectively) were used to heat a 1 cm section of one end of each channel at a current of 0.5 a for 10 seconds. we measured the melting point of the oil, water, and nacl-water mixture using a pal mythermpro melting point apparatus (mettler-toledo) and found it to be 25.5c. thus a 0.5 c gradient between 25.5 and 25c was established along each channel by using a digital multimeter and a 50-ohm resistor (variable resistance) in series with the two ends of the heater. in this figure, the isotherms (plotted as contours with colors) are colored by temperature, ranging from the freezing point of the deionized water in channel a (at 0c) to the freezing point of the 8% (m/m) nacl solution in channel b (at 5.09c). since the isotherms are colored by temperature, the effect of the overall heating of the chip is fairly negligible. as the temperature difference along the channel increases, the number of solid-liquid interfaces increases. note that as the temperature differences along the channel increase, the number of isotherms increases as well. the highest and lowest temperatures in the chip are at the solid-liquid interface locations, which are farther from the top of the chip than the locations of the isotherms. because the solid-liquid interfaces are located further from the top of the chip, the maximum temperature that the sample experiences will also be lower. this is true, even though the lowest temperature of the sample (the deionized water in channel a) is the highest temperature that the isotherms reach. 5ec8ef588b

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